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RESEARCH ARTICLE

Study of the Effect of some Antibiotics on Pseudomonas Aeruginosa Isolated from Wound and Burn Abscesses

Khansaa Basem Fadhil^{1*}, Ibrahim Omar Saeed², and Mahmood Khalaf Saleh³

^{1, 2} Department of Biology, College of Science, Tikrit University, Iraq

³ Department of Biology, College of Education for Pure Science, Tikrit University, Iraq

ARTICLE INFO	ABSTRACT							
Received: May 22, 2024	The study included the collection of 230 swabs from patients with wound							
Accepted: Jul 3, 2024	and burn infections from Salah al-Din Hospital from January to October, and a number of chemistry tests diagnosed developing colonies. 154							
	clinical isolates (96 burn isolates and 58 wound isolates) of the total							
Keywords	number of isolates could be diagnosed and the diagnosis was confirmed using the API 20E system. The isolates were found to have multiple							
Anaesthesia	antibiotic resistance as all isolates showed 100% resistance to Ampicillin							
Burns	and variable resistance to Norfloxacin, Aztreonam, Ceftriaxone, Ciprofloxacin and Trimethoprim (72%, 80%, 86%, 87% and 88%)							
Wounds	respectively, and showed sensitivity to Ofloxacin and Piperacillin (80							
Antibiotics	and 90%), respectively, and high sensitivity to Meropenem and Imipenem (100%).							
Pseudomonas Aeruginosa								

*Corresponding Author:

Khnsaa.bf@ntu.edu.iq

INTRODUCTION

Nosocomial infections are among the most complex health issues facing physicians worldwide. They are of great importance in public health issues nowadays despite the use of modern techniques in performing surgical operations and the good care shown by medical staff towards burns and wounds patients, causing an increase in human and economic damage, as well as an increase in treatment costs and an increase in the length of stay in the hospital [1], and [2]. Pseudomonas aeruginosa is one of the leading bacteria causing these infections [3]. Pseudomonas aeruginosa is a major cause of Pseudomonas aeruginosa is a major cause of burn infections that have claimed the lives of many burn patients and are a cause of high morbidity and mortality, These infections are caused by either endogenous pathogens (Endogenous), represented by the normal flora on the skin, intestines and respiratory tract of recumbents, or exogenous, such as transmission from medical staff, visitors, surgical instruments, air, water, food and floors [4]. [5] The indiscriminate use of antibiotics, including beta-lactam antibiotics, in hospitals and communities is the reason for the emergence of resistant strains and ineffective treatment and the emergence of strains characterized by multiple resistance to the antibiotic, which is a medical issue due to the difficulty of controlling diseases due to antibiotic resistance. Beta-lactam enzymes are at the forefront of the means used by

microorganisms to resist the action of beta-lactam antibiotics [6], and [7]. In addition, P. aeruginosa possesses an outer membrane that contains protein channels (Porins), and any alteration, modification, or reduction in the number of these channels leads to resistance to many antibiotics [8], and [9].

RESEARCH METHODOLOGY

Sample collection: A total of 230 samples were collected from inpatients and outpatients with burn and wound infections at Tikrit Teaching Hospital.

Diagnosis: Samples were cultured on developmental media and then re-cultured on selective media such as citarmide medium and the diagnosis was confirmed using API 20 Vitik.

Antibiotic susceptibility testing: The sensitivity of the isolates was studied by using antibiotic discs on an Acker-Muller-Hinton medium according to the method [10]

RESULTS AND DISCUSSION

Scientific studies indicate a disparity in the isolation rates of bacteria due to the difference in the source of isolation, the number of samples, the geographical location, the method of sterilization of wounds or burns, and the number of times sterilizations, as well as the common and indiscriminate use of antibiotics, which played a major role in the emergence of resistance to these bacteria [11].

The distribution of the isolated bacteria according to the isolation sources according to the final diagnostic results is shown in Table (1). It is clear from the table that P. aeruginosa was the most prevalent in burn infections (62%), followed by wound infections (38%). This indicates that P. aeruginosa is the main cause of burn abscesses. The results of P. aeruginosa were consistent with a study conducted in Iran by [5]. The percentage of P. aeruginosa was 73.1%, and the current study also agreed with the study [12] that P. aeruginosa was the most isolated bacterium in burn wounds.

Sensitivity of P. Aeruginosa to antibiotics

The sensitivity of Pseudomonas aeruginosa to antibiotics was tested in Table (2) as P. aeruginosa isolates showed high resistance to beta-lactam antibiotics. P. aeruginosa showed high resistance to beta-lactam antibiotics, as the resistance rate to Ampicillin was 100%. This resistance is due to a change in the structure of Penicillin Binding Proteins (PBPs) or the production of beta-lactam enzymes that can attack a wide range of beta-lactam antibiotics, The results of the study agreed with the findings of [13], while the isolates showed different resistance to Ceftriaxone, Ciprofloxacin, Norfloxacin, Aztreonam, and Trimethoprim.) 72%, 80%, 86%, 87%, and 88%, respectively. The results of the current study are in agreement with [14], and [15]. The reason for multiple resistance is attributed to the presence of resistance genes in clusters, which may be transferred together to the recipient cell by special DNA elements known as integrons located on the plasmid or bacterial chromosome, and the capture of genes that code for resistance can be restricted by the Recombination process, which enables integrons to attach to one or more gene cassette within its binding sites and thus form clusters of antibiotic resistance genes and beta-lactam antagonists [16]. While the majority of isolates showed sensitivity to Ofloxacin and Piperacillin by (80% and 90%), respectively, which is consistent with the findings of [17], and all isolates showed high sensitivity to Imipenem and Meropenem by 100%, the results agreed with the findings of [18], and [14], who found that P. aeruginosa is highly sensitive to Piperacillin. Carbapenems are very effective in treating infections caused by P. aeruginosa due to their permeability to the outer membrane of the bacteria and the increased resistance of Gram-negative bacteria has become one of the serious problems in Iraq and the world [12]. The reason for this is that they are broad-spectrum antibiotics against Grampositive and Gram-negative bacteria, in addition, these antibiotics are newly used, and their use is limited to hospitalized patients only because their administration must be mediated by the

physiological solution Normal saline intravenously for a period specified by the treating physician, and this is one of the reasons for the sensitivity of bacteria to them [19-20].

This study attempts to understand the importance of East Asia and the nature of conflicts and conflicts in the region, and how they have affected the formation of balances in the region.

REFERENCES

- A. Malik, S. E. Hasani, M. Shahid, H. M. Khan, and A. J. Ahmad, "Nosocomial Klebsiella infection in neonates in a tertiary care hospital: protein profile by SDS-page and klebocin typing as epidemiological markers," Indian J Med Microbiol, vol. 21, no. 2, pp. 82–86, 2003.
- M. J. Schwaber, S. E. Cosgrove, H. S. Gold, K. S. Kaye, and Y. Carmeli, "Fluoroquinolones protective against cephalosporin resistance in gram-negative nosocomial pathogens," Emerg Infect Dis, vol. 10, no. 1, p. 94, 2004.
- A. Jain and K. Singh, "Recent advances in the management of nosocomial infections," JK Science, vol. 9, no. 1, pp. 3–8, 2007.
- L. M. ; H. J. P. and K. D. A. Prescott, Microbiology. , 6th ed. U.S.A.: McGraw-Hill., 2005.
- A. R. R. Lari, R. Alaghehbandan, and L. Akhlaghi, "Burn wound infections and antimicrobial resistance in Tehran, Iran: an increasing problem," Ann Burns Fire Disasters, vol. 18, no. 2, p. 68, 2005.
- O. Ciofu, T. J. Beveridge, J. Kadurugamuwa, J. Walther-Rasmussen, and N. Høiby, "Chromosomal βlactamase is packaged into membrane vesicles and secreted from Pseudomonas aeruginosa," Journal of Antimicrobial Chemotherapy, vol. 45, no. 1, pp. 9–13, 2000.
- G. J. Tortora, B. R. Funke, and C. L. Case, Microbiology. Benjamin-Cummings Publishing Company, 1989.
- P. Lambert, "Mechanisms of antibiotic resistance in Pseudomonas aeruginosa.," J R Soc Med, vol. 95, no. Suppl 41, p. 22, 2002.
- D. M. Livermore, "Multiple mechanisms of antimicrobial resistance in Pseudomonas aeruginosa: our worst nightmare?," Clinical infectious diseases, vol. 34, no. 5, pp. 634–640, 2002.
- K. W. S. J. T. M. AW, "Antibiotic susceptibility testing by a standardized single disk method.," Am J Clin Pathol, vol. 4, no. 5325707., pp. 493–6, Apr. 1966.
- S.-H. Kang and M.-K. Kim, "Antibiotic sensitivity and resistance of bacteria from odontogenic maxillofacial abscesses," J Korean Assoc Oral Maxillofac Surg, vol. 45, no. 6, p. 324, 2019.
- F. D. Mahmoud, "Determination of the bioactivity of some plant extracts against molecularly characterised bacterial species isolated from different skin and environmental mixtures," Tikrit University, Iraq, 2022.
- M. B Khorsheed and S. S Zain Al Abdeen, "The frequency of Pseudomonas aeruginosa bacteria with some pathogenic bacteria in burns injuries and study their resistance to antibiotics," Kirkuk Journal of Science, vol. 12, no. 1, pp. 123–140, 2017.
- N. M. Khalaf, "Evaluation of the efficacy of some plant extracts on beta-lactamase-producing pathogenic bacteria isolated from skin abscesses," University of Diyala, Iraq, 2020.
- D. W. Khaled and B. A. Abdullah, "Antibiotic resistant infection of the bacterial group ESKAPE," 2018.
- Y. Khosravi, S. T. Tay, and J. Vadivelu, "Analysis of integrons and associated gene cassettes of metalloβ-lactamase-positive Pseudomonas aeruginosa in Malaysia," J Med Microbiol, vol. 60, no. 7, pp. 988–994, 2011.
- I. O. S. T. A. Z. Fanar Dawas Mahmood, " Detection of resistance bacteria isolated from skin infection to many antibiotics in several hospitals Mosul city," Tikrit University, Iraq, 2022.
- S. S. and A. Z. A. Zainul Abedin, "Investigation of induced betalactamases in Pseudomonas aeruginosa isolated from disease models," Kirkuk University Journal, Iraq, pp. 71–92, 2015.
- Jalal, B. J., & Alqaisi, M. R. M. (2024). Improving the production and quality of white button mushroom (Agaricus bisporus) by adding biochar and ash to the casing layer. Tikrit Journal for Agricultural Sciences, 24(1), 22–33. https://doi.org/10.25130/tjas.24.1.3.

Karim, K. K., & Abdulla, N. R. (2024). Use of various sources of calcium in the diets of broiler and its effects on carcass and some meat quality. Tikrit Journal for Agricultural Sciences, 24(1), 45–56. https://doi.org/10.25130/tjas.24.1.5.

	P. aeruginosa							
	Number Percentage							
Burn infections	96	62%						
Wound infections	58	38%						
Total	158	100%						

Table 1: Bacteria isolated	according to the	e sources of i	solation
Table 1. Ducteria isolatea	according to the	c sources or h	Joiation

Table 2: Percentage of resistance and	l sensitivity of Pseudomonas	aeruginosa
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Antibiotic	Sensitivity rate	Resistance rate
Ampicillin	%0	%100
Ceftriaxone	%14	%86
Trimethoprim	%12	%88
Aztreonam	%20	%80
Norfloxacin	%28	%72
Ciprofloxacin	%13	%87
Piperacillin	%90	%10
Ofloxacin	%80	%20
Meropenem	%100	%0

Imipenem	%100	%0

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2	APPA	-	3	ADO		4	PyrA	-	5	IARL	-	7	dCEL		9	BGAL	-
10	H2S	-	11	BNAG		12	AGLTp		13	dGLU	+	14	GGT	÷	15	OFF	•
17	BGLU		18	dMAL	+	19	dMAN	- 2	20	dMNE	÷	21	BXYL	-	22	BAlap	-
23	ProA	+	26	LIP	,+.,	27	PLE		29	TyrA	+	31	URE		32	dSOR	•
33	SAC	+	34	dTAG	7	35	dTRE	-	36	CIT	+	37	MNT	-	39	5KG	-
40	ILATk	+	41	AGLU	-	42	SUCT	÷	43	NAGA	-1	44	AGAL	-	45	PHOS	•
46	GlyA	-	47	ODC	-	48	LDC		53	IHISa	÷	56	CMT	+	57	BGUR	-
58	0129R	+	59	GGAA	-	61	IMLTa	+	62	ELLM	•	64	ILATa	+			

Figure 1: VITIK P. aeruginosa





Figure 2: API 20